PREDIdicting bacterial PATHogenicity on plant: PREDIPATH

Felipe LIRA¹, Gilles HUNAULT², Martial BRIAND¹, Perrine PORTIER¹, Claudine LANDES¹, and Marion FISCHER-LE SAUX¹



¹IRHS, INRA, Université d'Angers, Agrocampus-Ouest, SFR 4207 QuaSaV, 49071, Beaucouzé, France.

²Hémodynamique, Interaction Fibrose et Invasivité Tumorales Hépatiques Laboratory, Unité Propre de Recherche de l'Enseignement Supérieur 3859, Structure Fédérative de Recherche 4208, Bretagne Loire University, Angers, France.

Corresponding Author: felipelira3@gmail.com



E. tracheiphila BuffGH
Erwinia sp. ErVv1
Erwinia sp. 9145
E. oleae DAPP-PG531

Erwinia sp. OLMDLW33

persicina NBRC102418

pyrifoliae EpK1 15 pyrifoliae DSM12163

. amylovora LA092 . amylovora CTST01-1 . amylovora MAGFLF2

amylovora NHWL02-2

amylovora VTDMSF02

amylovora CFBP1232

amylovora NBRC1268 amylovora CFBP2585

. amýlovora 01SFR-BO . amylovora UPN527

amýlovora CFPB1430 amylovora ACW56400

pyrifoliae Ep1-96

Erwinia sp. Leaf53 Erwinia sp. B116

persicina B64

Backgrounds

Prediction of bacterial pathogenicity commonly relies on microbiological methods. **Comparative genomics** emerges as a efficient method for distinction and detection of genomics elements able to distinguish two or more classes of organisms (**pathogenic vs. non-pathogenic; commensal vs. free-living organisms**). Genes, genes clusters, and operons, are closely associated with the bacterial survival and spread. Most of them are exclusive and determinants to characterize bacterial groups. In order to facilitate the prediction of potential bacterial plant-pathogenicity of plant-associated bacteria, we propose the **PREDIPATH workflow**.

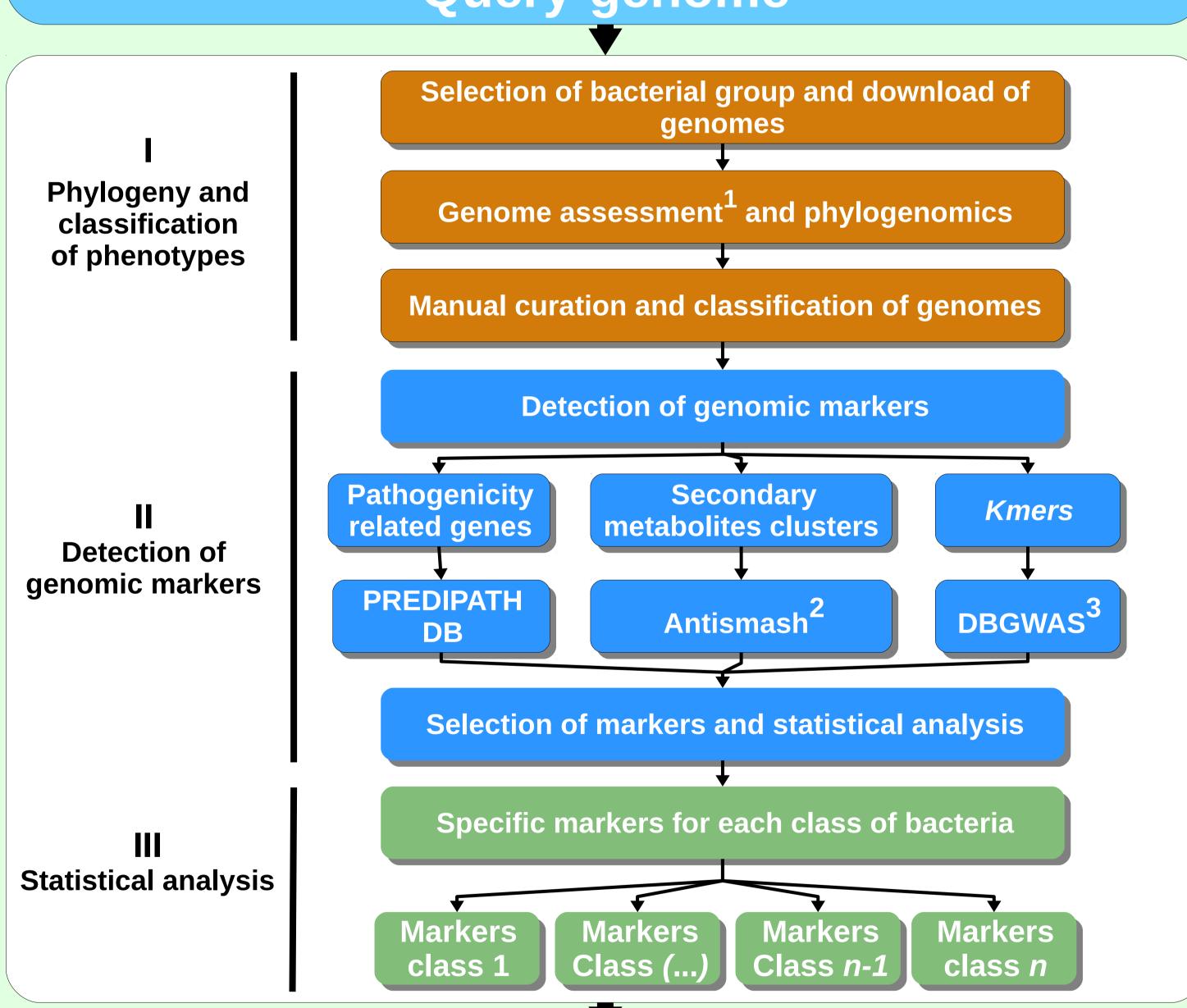
Objective

Creation of specific datasets of genes, clusters and sequence markers (*kmers*) to discriminate bacterial species based on their genomic sequences.

Development

The PREDIPATH methodology relies on the detection of genome-based markers and creation of specific datasets of markers enabling to identify potential pathogenic organisms based on their genomes. PREDIPATH pipeline was developed using Python programming language and external bioinformatics tools in the process. Our methodology for detection of markers is summarized in three major **Steps**:

Query genome



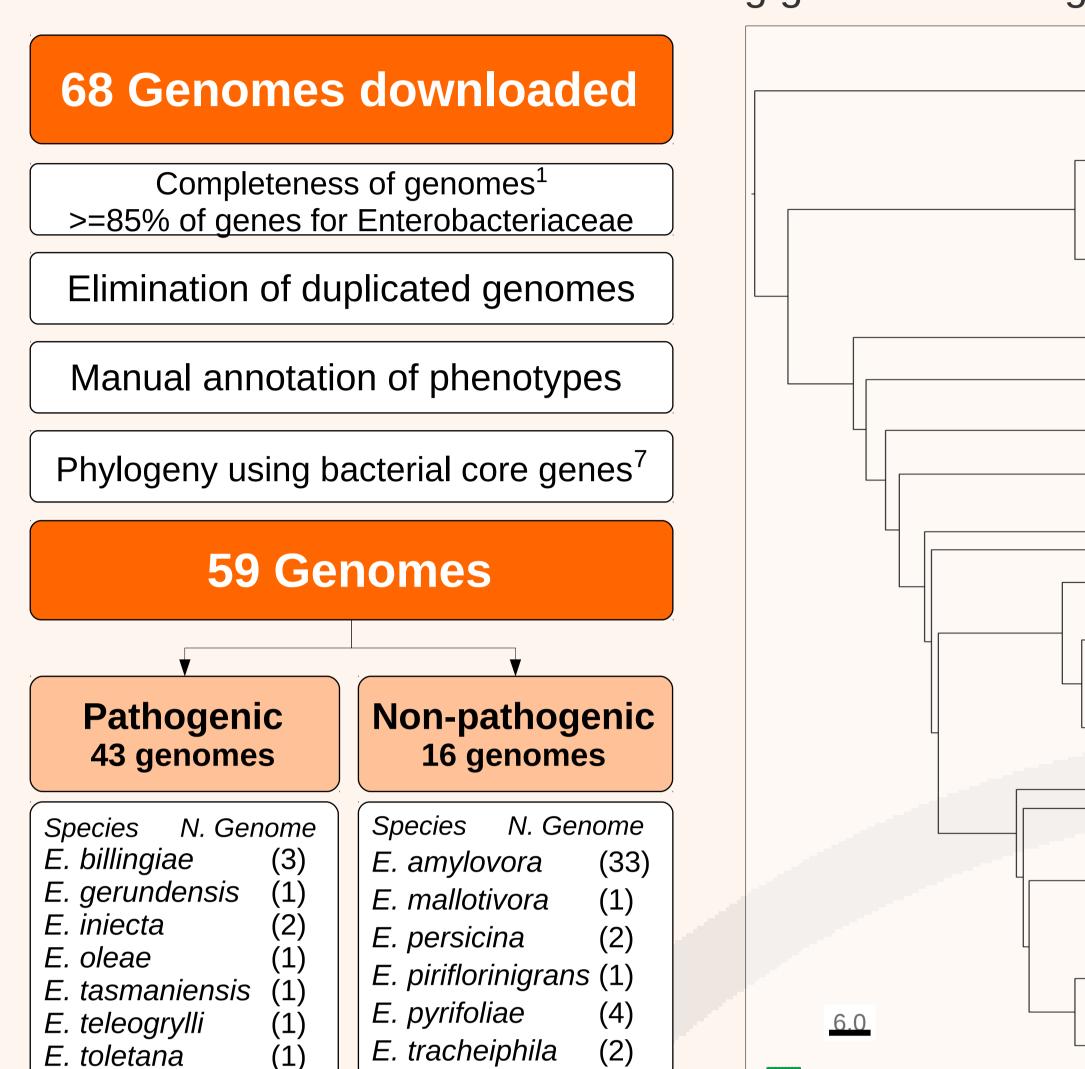
Bacterial classification based on their profile

Step I prioritizes the correct assignation of genomes in their classes and the correction of their nomenclature when needed. **Step II** consisted in to create a customized database to detect potential genes to be used as markers - **PREDIPATH Database** (*a priori* approach); the detection of differential secondary metabolites clusters, and small DNA fragments, such as *Kmers* exclusive for each class of organisms described in **Step I. Step III** gave support for the results obtained in **Step II**.

Benchmarking

PREDIPATH Database was compiled clustering the data from public repositories^{4,5,6} comprising a non-redundant dataset of sequences close-related with bacterial virulence and antimicrobial resistance.

PREDIPATH Workflow was tested using genomes from genus Erwinia.



After processing and detection of genes, secondary metabolites clusters, and kmers, simple and multiple binary logistic regressions were applied to identify specific markers.

Non-pathogenic

Pathogenic

	PREDIPATH DB	Sec.Metabolites Clusters	Kmers	
Total	14.248	24	1.143.473	Genomic elements
Detected	213	9	512	Total of markers detected
Non-pathogenic	-	-	51	Predictors* of classes
Pathogenic	5	9	12	*statistically significants

Simple binary logistic regression with PREDIPATH DB results were able to define a profile to predict the potential pathogenicity of plant-associated species:

- fur transcriptional repressor of iron-responsive genes
- hrpT type III secretion lipoprotein
- hrpF type III secretion protein
- hrpJ Hypersensitivity response secretion protein
- parE fluoroquinolones resistance gene
- A complete multiple binary logistic regression was able to predict the class using 9 variables only: thiopeptide, HSER, HSER arylpolyene, nrps, siderophore, terpene, butyrolactone, arylpolyene t1pks.
- Kmers exclusive to NP class were present from 19 to 100% of genome; exclusive kmers in class P were distributed between 7 to 53% of the genomes.

Conclusions

E. typographi

Erwinia sp.

- Phylogenetic distribution was not able to distinguish between pathogenic and non-pathogenic organisms in genus *Erwinia*.
- Our approach enable the compilation of a complete genomic datasets, composed by genes, clusters and kmers.
- Detection of exclusive markers by comparative genomics using the **PREDIPATH** workflow allowed the creation of exclusive datasets of predictors to diagnostic potential pathogenicity of plant-associated bacteria.

















^{1.}RM Waterhouse, M Seppey, FA Simão, M Manni, P Ioannidis, G Klioutchnikov, E V Kriventseva, EM Zdobnov, BUSCO Applications from Quality Assessments to Gene Prediction and Phylogenomics, Molecular Biology and Evolution, Volume 35, Issue 3, March 2018, Pages 543–548

^{2.}Blin K, Wolf T, Chevrette MG, et al. antiSMASH 4.0-improvements in chemistry prediction and gene cluster boundary identification. *Nucleic Acids Res.* 2017;45(W1):W36-W41.

^{3.}Jaillard M, Lima L, Tournoud M, Mahé P, *et al.* (2018) A fast and agnostic method for bacterial genomewide association studies: Bridging the gap between kmers and genetic events. *PLoS Genetics* 14(11).

^{4.}Chen LH, Yang J, Yu J, Yao ZJ, Sun LL, Shen Y and Jin Q, 2005. VFDB: a reference database for bacterial virulence factors. *Nucleic Acids Res.* 36 (Database issue):D539-D542.

^{5.}Jia B, Raphenya AR, Alcock B, et al. CARD 2017: expansion and model-centric curation of the comprehensive antibiotic resistance database. *Nucleic Acids Res.* 2016;45(D1):D566-D573.

^{6.}Pal, C., Bengtsson-Palme, J., Rensing, C., Kristiansson, E., Larsson, DGJ. (2014) BacMet: antibacterial biocide and metal resistance genes database, *Nucleic Acids Res.*, 42, D737-D743